

Role of peptide modifications in differentially modulating multiple components of a rhythmic neuromuscular system in the American lobster,

may undergo further posttranslational modification (PTMs) before becoming fully bioactive. These modifications regulate how peptides interact with the cellular environment, ensuring they bind proper bioactivity. When bound to their receptor, peptides can alter neuronal function and elicit unique behaviors.

The goal of this project was to further elucidate the molecular underpinnings of peptidergic modulation through physiological experiments combined with transcriptomics and molecular biology. The CPG that drives the rhythmic contractions of the lobster heart, the cardiac ganglion (CG), serves as an extremely simple network composed of only nine neurons that produces quantifiable rhythmic output. One such modulator is myosuppressin (pQDLDHVFLRFamide). Previous research from the Dickinson lab (Stevens, et al., 2009) has shown that, when applied to whole hearts, myosuppressin causes an immediate decrease in contraction frequency and contraction amplitude, followed by a large increase in contraction amplitude. By applying myosuppressin to isolated CGs and externally stimulated muscle cells, it was found that myosuppressin acts both centrally, on the neurons themselves, and peripherally, at the neuromuscular junction. It was found that myosuppressin alters a number of the burst characteristics of the CGs, as predicted

by three of which exist in the CG. In addition, it has been shown that myosuppressin exists endogenously in the lobster with two PTMs: cyclization of the (N)-terminal glutamine and amidation at the (C)-terminus. It may also exist in forms that lack certain modifications, which may be differentially bioactive. My previous research tested three isoforms of myosuppressin on whole hearts of lobsters; one isoform with both modifications, one isoform lacking the cyclization, and one isoform lacking amidation. I found that both the fully modified isoform, and the cyclized isoform were able to elicit a decrease in frequency and a decrease in amplitude followed by a large increase. In contrast, the non-amidated isoform was able to elicit a smaller decrease in frequency and amplitude but was not able to elicit an increase in amplitude. This summer I investigated whether the differentially modified isoforms of myosuppressin were able to respond in the periphery of the system, in the absence of neuronal input. To do this, I dissected out the cells of the CG and provided manual stimulation at a nerve ending to evoke contractions of the muscle. In this experiment, the fully modified isoform of myosuppressin and the cyclized form caused an increase in the amplitude of evoked contractions, whereas the non-amidated form caused no change in the amplitude of contractions, suggesting the non-amidated isoform is unable to bind either at the neuromuscular junction or muscle.

To further investigate the binding capabilities of myosuppressin, I began to characterize the expression of the predicted receptor sequences in the neural tissue of the lobster. Using real time PCR, four predicted myosuppressin sequences were identified in the CG, two of which were also identified in the muscle. One receptor was identified in

